IOTest®

CD5-PE

REF A07753
100 tests; 2 mL
20 µL / test

BECKMAN COULTER™

IOTest
Conjugated Antibody

IVD CE

ENGLISH

Specifications	
Specificity	CD5
Clone	BL1a
Hybridoma	SP2/0-Ag14 x Balb/c
Immunogen	Lymphocytes from the human thoracic duct
Immunoglobulin	IgG2a
Species	Mouse
Source	Ascites
Purification	Protein A affinity chromatography
Fluorochrome	R Phycoerythrin (PE)
λ excitation	488 nm
Emission peak	575 nm
Buffer	Buffer (PBS pH 7.2) plus 2 mg / mL BSA and 0.1% NaN_3

USE

This fluorochrome-conjugated antibody permits the identification and numeration of cell populations expressing the CD5 antigen present in human biological samples using flow cytometry.

PRINCIPLE

This test is based on the ability of specific monoclonal antibodies to bind to the antigenic determinants expressed by leucocytes.
Specific staining of the leucocytes is performed by incubating the sample with the IOTest reagent. The red cells are then removed by lysis and the leucocytes, which are unaffected by this process, are analyzed by flow cytometry.
The flow cytometer measures light diffusion and the fluorescence of cells. It makes possible the delimitation of the population of interest within the electronic window defined on a histogram, which correlates the orthogonal diffusion of light ("Side Scatter" or SS) and the diffusion of narrow-angle light (“Forward Scatter” or FS). Other histograms combining two of the different parameters available on the cytometer, can be used as supports in the electronic gating stage depending on the application chosen by the user.
The fluorescence of the delimited cells is analyzed in order to distinguish the positively-stained events from the unstained ones. The results are expressed as a percentage of positive events in relation to all the events acquired by the gating.

EXAMPLES OF CLINICAL APPLICATIONS

The CD5 molecule is a useful marker for the identification of certain lymphoproliferative syndromes when used within an extended immunophenotyping panel. Thus, the co-expression of CD5 and CD19 antigens enables chronic B cell lymphoid leukaemias (B-CLL : $CD5^+$ $CD19^+$) to be differentiated from T cell lymphoproliferative syndromes ($CD5^+$ $CD19^-$) (1, 2).
The simultaneous analysis of CD5, CD19 and CD23 antigens enables the different B cell proliferations to be more accurately distinguished, such as chronic B cell lymphoid leukaemias and small cell lymphomas, which are $CD5^+$ $CD23^+CD19^+$ and mantle lymphomas, which are $CD5^+CD23^-$ $CD19^+$ (1 – 6).
The simultaneous analysis of CD5, CD19 and CD10 antigens helps the characterization of different B-cell neoplasias (7), such as chronic B-cell lymphoid leukaemias and small-cell lymphomas, which are $CD5^+CD10^-$ $CD19^+$ and follicular lymphomas, which are $CD5^-$ $CD10^+$ $CD19^+$ (2, 3). This analysis also permits B acute lymphoblastic leukaemias (B-ALL) and pre-B acute lymphoblastic leukaemias which are $CD5^-$ $CD10^+CD19^+$, to be differentiated from prolymphocytic leukaemias which are $CD5^-$ $CD10^-$ $CD19^+$ (2).

STORAGE AND STABILITY

The conjugated liquid forms must be kept between 2°C and 8°C and protected from light, before and after the vial has been opened.
Stability of closed vial: see expiry date on vial.
Stability of opened vial: the reagent is stable for 90 days.

PRECAUTIONS

1. Do not use the reagent beyond the expiry date.
2. Do not freeze.
3. Let it come to room temperature (18–25°C) before use.
4. Minimize exposure to light.
5. Avoid microbial contamination of the reagents, or false results may occur.
6. Antibody solutions containing sodium azide (NaN_3) should be handled with care. Do not take internally and avoid all contact with the skin, mucosa and eyes.
   Moreover, in an acid medium, sodium azide can form the potentially dangerous hydrazoic acid. If it needs to be disposed of, it is recommended that the reagent be diluted in a large volume of water before pouring it into the drainage system so as to avoid the accumulation of sodium azide in metal pipes and to prevent the risk of explosion.
7. All blood samples must be considered as potentially infectious and must be handled with care (in particular: the wearing of protective gloves, gowns and goggles is recommended).
8. Never pipette by mouth and avoid all contact of the samples with the skin, mucosa and eyes.
9. Blood tubes and disposable material used for handling should be disposed of in ad hoc containers intended for incineration.

SAMPLES

Venous blood or bone marrow samples must be taken using sterile tubes containing an EDTA salt as the anticoagulant. The use of other anticoagulants is not recommended.
The samples should be kept at room temperature (18–25°C) and not shaken. The samples should be homogenized by gentle agitation prior to taking the test sample.
The samples must be analyzed within 24 hours of taking them.

METHODOLOGY

NECESSARY MATERIAL NOT SUPPLIED

- Sampling tubes and material necessary for sampling.
- Automatic pipettes with disposable tips for 20, 100 and 500 µL.
- Plastic haemolysis tubes.
- Calibration beads: Flow-Set™ Fluorospheres (Ref.6607007).
- Red cell lysis reagent with washing stage after lysis. For example: VersaLyse (Ref. A09777).
- Leucocyte fixation reagent. For example : IOTest 3 Fixative Solution (Ref. A07800).
- Negative control: isotypic control IgG2a-PE mouse.
- Buffer (PBS: 0.01 M sodium phosphate; 0.145 M sodium chloride; pH 7.2).
- Centrifuge.
- Automatic agitator (Vortex type).
- Flow cytometer.

PROCEDURE

NOTE: The procedure below is valid for standard applications. Sample and/or VersaLyse volumes for certain Beckman Coulter applications may be different. If such is the case, follow the instructions on the application’s technical leaflet.
For each sample analyzed, in addition to the test tube, one control tube is required in which the cells are mixed in the presence of an isotypic control.

1. Add 20 µL of specific IOTest conjugated antibody to each test tube, and to each control tube, the necessary amount of the isotypic control.
2. Add 100 µL of the test sample to both tubes. Vortex the tubes gently.
3. Incubate for 15 to 20 minutes at room temperature (18–25°C), protected from light.
4. Then perform lysis of the red cells, if necessary, by following the recommendations of the lysis reagent used.
   As an example, if you wish to use VersaLyse (Ref. A09777), refer to the leaflet and follow preferably the procedure called “with concomitant fixation”, which consists of adding 1 mL of the “Fix-and-Lyse” mixture prepared extemporaneously. Vortex immediately for one second and incubate for 10 minutes at room temperature, protected from light.
   If the sample does not contain red cells, add 2 mL of PBS.
5. Centrifuge for 5 minutes at 150 x g at room temperature.
6. Remove the supernatant by aspiration.
7. Resuspend the cell pellet using 3 mL of PBS.
8. Repeat step 5.
9. Remove the supernatant by aspiration and resuspend the cell pellet using:

– 0.5 mL or 1 mL of PBS plus 0.1% of formaldehyde if the preparations are to be kept for more than 2 hours and less than 24 hours. (A 0.1% formaldehyde PBS can be obtained by diluting 12.5 µL of the IOTest 3

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Fixative Solution (Ref. A07800) at its 10X concentration in 1 mL of PBS).
- 0.5 mL or 1 mL of PBS without formaldehyde, if the preparations are to be analyzed within 2 hours.

NOTE: In all cases, keep the preparations between 2°C and 8°C and protected from light

PERFORMANCE

SPECIFICITY

The CD5 molecule is expressed at the surface of mature T lymphocytes, by the majority of thymocytes and by a sub-population of B lymphocytes (8 – 10). Its expression is not found in granulocytes, monocytes and platelets (10).
The BL1a mAb was assigned to CD5 during the 3rd HLDA Workshop on Human Leucocyte Differentiation Antigens, held in Oxford, England, in 1986 (Code WS: 520, Section T) (8, 9).

LINEARITY

To test the linearity of staining of this reagent, a positive cell line (MOLT4) and a negative cell line (RPMI) were mixed in different proportions with a constant final number of cells, so that the positive line / negative line ratio of the mixture ranged from 0% to 100%.
Aliquots were stained using the procedure described above and linear regression between the expected values and the observed values was calculated. The parameters of the equation of the linear regression may be used to determine the linearity as well as the range of measurement.

Specificity	Linear regression	Linearity (R^2)	Range (%)
CD5	Y = 1.00 X + 0.09	0.9998	1 – 99

EXPECTED VALUES

Each laboratory must compile a list of reference values based upon a group of healthy donors from the local population. This must be done by taking age, sex and ethnic group into account, as well as any other potential regional differences.
In our laboratories, the whole blood of 50 healthy adults was treated using the IOTest 3 CD5-FITC /CD23-PE / CD19-ECD reagent (Ref. A07710). The results obtained for the percentage of the positive events of interest ($CD5^+$ lymphocytes) are given in the table below:

Lymphocytes	Number	Mean %	SD	CV (%)
$CD5^+$	50	73	7.7	10.5

INTRA-LABORATORY REPRODUCIBILITY

On the same day and using the same cytometer, 12 measurements of the percentage of staining were carried out on a target population (lymphocytes). The results obtained are summarized in the following table:

Positive Target	Number	Mean %	SD	CV (%)
$CD5^+$ Lymphocytes	12	73	2.7	3.7

INTER-LABORATORY REPRODUCIBILITY

On the same day and for the same positive target (lymphocytes), 12 measurements of the percentage of stained cells were carried out by two technicians and the preparations analyzed using two different cytometers. The results obtained are summarized in the following tables:

Cytometer n° 1:

Positive Target	Number	Mean (%)	SD	CV (%)
$CD5^+$ Lymphocytes	12	73	2.7	3.7

Cytometer n° 2:

Positive Target	Number	Mean (%)	SD	CV (%)
$CD5^+$ Lymphocytes	12	76	0.7	1.0

LIMITATIONS OF THE TECHNIQUE

1. Flow cytometry may produce false results if the cytometer has not been aligned perfectly, if fluorescence leaks have not been correctly compensated for and if the regions have not been carefully positioned.
2. It is preferable to use a lysis technique with washing as this reagent has not been optimized for "without washing" lysis techniques.
3. Accurate and reproducible results will be obtained as long as the procedures used are in accordance with the technical insert leaflet and compatible with good laboratory practices.
4. The conjugated antibody of this reagent is calibrated so as to offer the best specific signal / non-specific signal ratio. Therefore, it is important to adhere to the reagent volume / number of cells ratio in every test.
5. In the case of a hyperleucocytosis, dilute the blood in PBS so as to obtain a value of approximately 5 x 10^9 leucocytes/L.
6. In certain disease states, such as severe renal failure or haemoglobinopathies, lysis of red cells may be slow, incomplete or even impossible. In this case, it is recommended to isolate mononucleated cells using a density gradient (Ficoll, for example) prior to staining.

MISCELLANEOUS

See the Appendix for examples and references.

TRADEMARKS

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APPENDIX TO REF A07753

EXAMPLES

The graphs below are monoparametric representations (Count vs. Fluorescence Intensity) of lyzed normal whole blood sample. Staining is with IOTest CD5-PE Conjugated Antibody (Ref. A07753). Gate is on lymphocytes. A mouse PE-conjugated IgG2a isotypic control is shown in light.

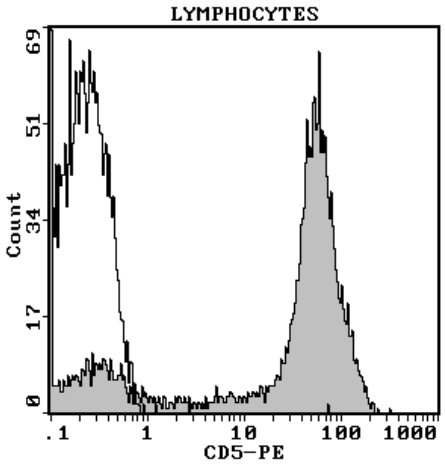

Figure 1: acquisition and analysis are performed with a COULTER® EPICS® XL™ flow cytometer equipped with System II™ software.

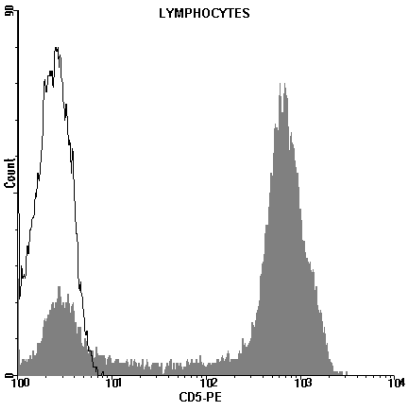

Figure 2: acquisition is with a Becton Dickinson FACScan™ flow cytometer.

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